Recent Progress Toward Hydrogen Medicine: Potential of Molecular Hydrogen for Preventive and Therapeutic Applications

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Abstract: Persistent oxidative stress is one of the major causes of most lifestyle-related diseases, cancer and the aging process. Acute oxidative stress directly causes serious damage to tissues. Despite the clinical importance of oxidative damage, antioxidants have been of limited therapeutic success. We have proposed that molecular hydrogen (H₂) has potential as a "novel" antioxidant in preventive and therapeutic applications [Ohsawa *et al.*, Nat Med. 2007: 13; 688-94]. H₂ has a number of advantages as a potential antioxidant: H₂ rapidly diffuses into tissues and cells, and it is mild enough neither to disturb metabolic redox reactions nor to affect reactive oxygen species (ROS) that function in cell signaling, thereby, there should be little adverse effects of consuming H₂. There are several methods to ingest or consume H₂, including inhaling hydrogen gas, drinking H₂-dissolved water (hydrogen water), taking a hydrogen bath, injecting H₂-dissolved saline (hydrogen saline), dropping hydrogen saline onto the eye, and increasing the production of intestinal H₂ by bacteria. Since the publication of the first H₂ paper in *Nature Medicine* in 2007, the biological effects of H₂ have been confirmed by the publication of more than 38 diseases, physiological states and clinical tests in leading biological/medical journals, and several groups have started clinical examinations. Moreover, H₂ shows not only effects against oxidative stress, but also various anti-inflammatory and antiallergic effects. H₂ regulates various gene expressions and protein-phosphorylations, though the molecular mechanisms underlying the marked effects of very small amounts of H₂ remain elusive.

Keywords: Anti-inflammation, antioxidant, hydrogen medicine, medical gas, mitochondria, oxidative stress, ischemia-reperfusion, ROS.

1. INTRODUCTION

Oxidative stress arises from the strong cellular oxidizing potential of excess reactive oxygen species (ROS) [1]. Acute oxidative stress arises from a variety of situations, including ischemia reperfusion [2]. Persistent oxidative stress is widely accepted as one of the causes of most lifestyle-related diseases, cancer and the aging process [3-7]; however, many antioxidant supplements could not prevent cancer, myocardial farction and atherosclerosis, but rather conversely increase mortality [8-11]; thus, it is very important to be aware of side effects when developing an effective antioxidant for the prevention of oxidative stress-related diseases.

We found that molecular hydrogen (H_2) has roles as a "novel" antioxidant in preventive and therapeutic applications [12]. H_2 has advantages as a potential antioxidant without adverse effects: it is mild enough neither to disturb metabolic redox reactions nor to affect ROS, which function in cell signaling [13-15] and has favorable distribution characteristics in its own physical ability to penetrate biomembranes and diffuse through barriers into cellular components.

Here, we review the recent progress toward therapeutic and preventive applications of H_2 in widespread fields.

2. ROS AS ONE OF THE MAJOR CAUSES OF ACUTE AND CHRONIC DISEASES

2.1. Persistent Oxidative Stress

ROS are generated inside the body throughout our daily lives, such as during hard exercise, smoking, exposure to ultraviolet rays or air pollution, aging, physical or psychological stress, and so on [16-19]. Inside every aerobic organism, ROS are generated when breathing consumes oxygen.

As the first step in generating persistent ROS, the majority of superoxide anion radicals ($\bullet O_2$) are generated in mitochondria by electron leakage from the electron transport chain [3, 7 20, 21].

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Superoxide dismutase converts to hydrogen peroxide (H_2O_2) , which is metabolized by glutathione peroxidase or catalase to generate water (H_2O) . Highly reactive hydroxyl radicals (\bullet OH) are generated from H_2O_2 via the Fenton or Weise reaction in the presence of catalytically active metals, such as Fe^{2+} and Cu^+ [22]; therefore, manipulation of the genes involved in anti-oxidation prolonged the lifespan or prevented disease models [23-27].

These ROSs are generated under the condition of excessively high membrane potential to leak electrons from the electron transport chain [28]. In fact, uncoupling proteins control the membrane potential to suppress the production of ROS and then consequently to repress diabetes [29-31].

Mitochondrial aldehyde hydydrogenase 2 (ALDH2) functions as a protector against oxidative stress by detoxifying cytotoxic aldehydes, such as 4-hydroxy-2-nonenal [4, 5, 32]. Thus, a defect of ALDH2 sufficiently induces phenotypes of age-dependent dementia by accumulating such cytotoxic aldehydes [32]. Paradoxically, such aldehydes stimulate protective systems against oxidative stress [33]. Thus, oxidative stress has two faces, to damage tissues and to enhance protective systems.

2.2. Acute Oxidative Stress

Acute oxidative stress arises from various different situations: inflammation, cardiac or cerebral infarction, organ transplantation, heavy exercise, cessation of operative bleeding, and others [2, 34, 35]. In many cases, ischemia reperfusion is a critical cause to raise acute oxidative stress. In myocardial infarction, the accelerated generation of ROS by reperfusion of the ischemic myocardium is a potential mediator of reperfusion injury [36-39]. During myocardial reperfusion, $\bullet O_2^-$ is generated within the injured mitochondria via electron leakage from the electron transport chain. $\bullet O_2^-$ converts to H_2O_2 , and highly reactive $\bullet OH$ is generated from H_2O_2 as mentioned [22, 40].

These ROS mediate myocardial injury by inducing mitochondrial permeability transition pore (PTP) opening, causing a loss of mitochondrial membrane potential, and leading to mitochondrial swelling with membrane rupture [41]. Many attempts have been made to inhibit ROS production to limit the extent of reperfusion

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injury. The administration of ROS scavengers at the time of reperfusion has produced conflicting results that can be partially explained by the dual role of ROS in ischemia-reperfusion hearts [42, 43]. The majority of detrimental effects associated with lethal reperfusion injury are attributed to •OH. By comparison, •O₂ and H₂O₂ have less oxidative energy and, paradoxically, are implicated as crucial signaling components in the establishment of tolerance to oxidative stress [44, 45]. Thus, cytotoxic radicals such as •OH must be neutralized without compromising the essential biological activities of other ROS, including NO• [46, 47].

3. CHARACTERISTICS OF MOLECULAR HYDROGEN

We found that H_2 functions as a mild but effective antioxidant [12]. Hydrogen is the most abundant element in the universe, constituting nearly 75% of the universe's mass; however, hydrogen is absent on the earth in its monoatomic form and is present in water and organic or inorganic compounds. Hydrogen gas, with the molecular formula H_2 , is a colorless, odorless, tasteless and highly combustible diatomic gas. The earth's atmosphere contains less than 1 part per million of hydrogen gas [48].

 $\rm H_2$ is rather less active and behaves as an inert gas in the absence of catalysts or at body temperature. $\rm H_2$ does not react with most compounds, including oxygen gas at room temperature. Hydrogen gas is flammable only at temperature higher than 527°C, and explodes by a rapid chain reaction with oxygen only in the explosive range of the $\rm H_2$ concentration (4 - 75%, vol/vol).

H₂ can be dissolved in water up to 0.8 mM (1.6 ppm, wt/vol) under atmospheric pressure, and rapidly H₂ penetrates the glass and plastic walls of any vessels, while aluminum containers are able to retain hydrogen gas for a long time.

4. SCAVENGING EFFECTS ON HYDROXYL RADICALS IN CULTURED CELLS

4.1. Scavenging •OH, but Not •O₂, H₂O₂ and NO in Cultured Cells

 H_2 scavenges •OH, but not •O₂, H_2O_2 and NO in cultured cells. H₂ was dissolved in culture medium under high pressure of hydrogen gas or by simply bubbling with hydrogen gas. The medium was combined with O_2 -saturated medium at the ratio of 8 : 2 (H_2 : O_2). Hydrogen and oxygen concentrations and pH were monitored with each specific electrode. Cultured cells were treated with a mitochondrial respiratory complex III inhibitor, antimycin, A to induce excess •O₂ production. Following such treatment, •O₂ was rapidly converted to H₂O₂ and then •OH. The addition of antimycin A actually increased levels of $\bullet O_2^-$ and H_2O_2 inside cells; however, H_2 dissolved in culture medium did not change their levels. Additionally, H2 did not decrease the steady-state level of NO in cells. In contrast, H₂ treatment significantly decreased levels of •OH, as judged by the decrease in the fluorescent signal of hydroxyphenyl fluorescein (HPF) [49] and in the spin trap signals. Notably, H₂ decreased •OH levels even in the nuclear region [12].

After antimycin A treatment, H_2 prevented the decline of the mitochondrial membrane potential. This suggested that H_2 protected mitochondria from \bullet OH. Along with this protective effect, H_2 also prevented a decrease in the cellular level of ATP synthesized in mitochondria. The fact that H_2 protected mitochondria and nuclear DNA provided evidence that H_2 penetrated most membranes and diffused into organelles. Consequently, H_2 protected cultured cells against oxidative stress [12].

4.2. Other Effects Shown by Using Culture Systems

H₂ dissolved in medium protected cultured auditory hair cells from free radicals [50] and is suggested to decrease •OH, as judged by the decrease in HPF fluorescence in vestibular tissue [51].

•OH causes most ionizing radiation-induced cellular damage. H₂ exhibited protective effects against radiation-induced damage in

cultured cells and mice [52]. Cosmic radiation is known to induce DNA and lipid damage associated with increased oxidative stress and remains a major concern in space travel. It is expected that space mission activities will increase in coming years both in number and duration. It is therefore important to estimate and prevent the risks encountered by astronauts due to oxidative stress prior to developing clinical symptoms of disease. Schoenfeld *et al.* hypothesized that H₂ administration to astronauts by either inhalation or drinking hydrogen water may potentially yield a novel and feasible preventative/therapeutic strategy to prevent radiation-induced adverse events [53].

On the other hand, H₂ treatment prolonged the replicable lifespan of bone marrow multipotential stromal cells in vitro while preserving differentiation and paracrine potentials. Cell therapy with bone marrow multipotential stromal cells/mesenchymal stem cells represents a promising approach in the field of regenerative medicine. Low frequency of mesenchymal stem cells in adult bone marrow necessitates ex vivo expansion of mesenchymal stem cells after harvest; however, such manipulation causes cellular senescence with loss of differentiation, proliferative, and therapeutic potentials of mesenchymal stem cells. As oxidative stress is one of the key insults promoting cell senescence in vivo as well as in vitro, H₂ prevented the senescent process during mesenchymal stem cell expansion. Notably, 3% hydrogen gas treatment did not decrease •OH, protein carbonyl, and 8-hydroxydeoxyguanosine, suggesting that scavenging •OH might not be responsible for these effects of hydrogen gas in this study [54].

5. ADVANTAGES OF HYDROGEN

5.1. Rapid Diffusion

H₂ has a number of advantages as a potential antioxidant. First, it has favorable distribution characteristics with its own physical ability to penetrate biomembranes and diffuse into the cytosol.

Excessive oxidative damage is a major factor because the mitochondrial respiratory chain is a significant source of damaging reactive oxygen species; however, despite the clinical importance of mitochondrial oxidative damage, antioxidants have been of limited therapeutic success. This may be because antioxidants are not selectively taken up by mitochondria [55-57]. As H₂ effectively reaches the nucleus and mitochondria, the protection of nuclear DNA and mitochondria suggests preventive effects on lifestyle-related diseases, cancer and the aging process [12]. Moreover, H₂ passes through the blood brain barrier, although most antioxidant compounds cannot; this is also an advantage of H₂.

Monitoring H₂ concentration inside various tissues can prove gaseous diffusion [58].

5.2. No Direct Elimination of Functionally Important ROS

Despite their cytotoxic effects, low concentrations of ROS, such as $\bullet O_2^-$ and H_2O_2 , function as signaling molecules and regulate apoptosis, cell proliferation, and differentiation [14, 15]. As mentioned, unexpectedly and notably, recent studies have suggested that excessive antioxidants increased mortality and rates of cancer [9, 11, 59-62] because they may interfere with some essential defensive mechanisms [13, 60, 63-67]. At higher concentrations, H_2O_2 is converted to hypochlorous acid by myeloperoxidase to defend against bacterial invasion [68]. Additionally, NO functions as a neurotransmitter and is essential for the dilation of blood vessels [69].

Since H_2 reduces \bullet OH but does not affect \bullet O₂ and H_2 O₂ having physiological roles [12], we propose that the adverse effects of H_2 are very small compared to other antioxidants.

5.3. No toxicity Even at Higher Concentration.

Several medical gasses are expected to provide more effective therapeutic interventions and preventive medicine despite their

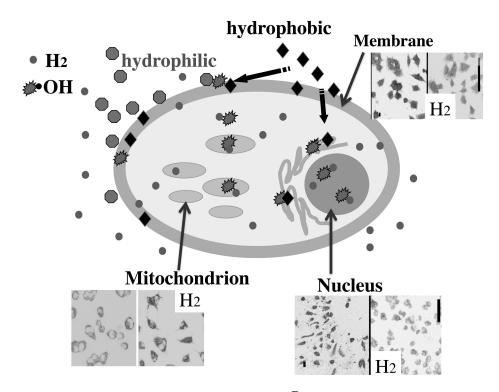


Fig. (1). Illustration of gaseous diffusion of H₂ in a cell. Most hydrophilic compounds () retain at membranes and cannot reach the cytosole, whereas most hydrophic ones (\P) cannot penetrate biomembranes in the absence of specific carriers. In contrast, H_2 (\P) can rapidly distribute into cytosol and organelles. PC12 cells were placed in culture media containing H₂ (0.6 mM) and O₂ (0.24 mM), and then oxidative stress was induced by adding antimycin A (10 µg/mL), an inhibitor of the electron transport chain of mitochondria, and maintained for 1 day. Two markers of oxidative stress were detected by immunostaining with anti-8-hydroxy-Guanine (Nucleus) and anti-4-hydroxy-2-nonenal (Membrane). Thirty minutes after adding antimycin A with or without H2, 100 nM tetramethylrhodamine methyl ester (TMRM), a fluorescent detector of the membrane potential of mitochondrion, were added, incubated for 10 min, and cells were imaged with a laser scanning confocal microscope. These results indicate that H₂ reach the nucleus and mitochondria and protects them.

severe toxicity. Gas inhalation as disease therapy has received recent interest [70]. In past decades, there has been extraordinary, rapid growth in our knowledge of gaseous molecules, including nitric oxide (NO), carbon monoxide (CO), and hydrogen sulfide (H₂S), which have been known to play important roles in biological systems [71, 72].

In pre-clinical experimental models of disease, including ischemia-reperfusion injury, the inhalation of exogenous CO or H₂S has produced a favorable outcome for most vital organs [73-76]. In particular, NO has been approved as a therapeutic agent in clinical practice [77]. The inherent toxicity of these gasses must be investigated for gas inhalation to be considered an effective therapeutic strategy because these gasses are highly toxic at considerable concentrations. Additionally, NO enhances oxidative stress via the reaction with $\bullet O_2^-$ by the production of highly oxidative peroxynitrite (NO + \bullet O₂⁻ \rightarrow ONOO⁻). It is unknown if the therapeutically effective threshold for CO or H₂S can be attained locally in target organs without delivering a potentially toxic level of the gasses via the lungs.

In contrast, H₂ has more advantages from the aspect of toxicity: H₂ has no cytotoxicity even at high concentration [78-81]. Furthermore, safety standards have been established for high concentrations of hydrogen gas for inhalation since high pressure hydrogen gas is used in deep diving gas mixes to prevent decompression sickness and arterial gas thrombi [81]. The safety of H₂ for humans is demonstrated by its application in Hydreliox, an exotic, breathing gas mixture of 49% H₂, 50% helium and 1% O₂, which is used to prevent decompression sickness and nitrogen narcosis during very deep technical diving [78-81].

6. METHODS OF INGEST HYDROGEN I: INHALATION OF HYDROGEN GAS

6.1. Inhalation of Hydrogen Gas

Inhalation of hydrogen gas is a straightforward therapeutic method. Hydrogen gas can be inhaled by delivering hydrogen gas through a ventilator circuit, facemask or nasal cannula. Since inhaled hydrogen gas acts more rapidly, it may be suitable for defense against acute oxidative stress. In particular, inhalation of gas does not affect blood pressure [12]; blood pressure increased by infusion may cause serious obstacles during the treatment of myocardial infarction. Hydrogen gas poses no risk of explosion in air and in pure oxygen when present at concentrations < 4%, as mentioned earlier; however, safety could be a concern and the desired concentration of H₂ must be monitored and maintained with an approved and commercially available tool.

Rats inhaled hydrogen gas in a mix of nitrous oxide (N₂O) (for anesthesia), O₂, and N₂. The inhalation of H₂ actually increased H₂ dissolved in arterial blood depending upon the hydrogen gas concentrations, and H2 levels in venous blood were lower than in arterial blood; the different level between arterial and venous blood indicates the amount of H_2 incorporated into tissues [12].

6.2. Direct Demonstration of Rapid Diffusion of Hydrogen as a Medical Gas

Gasses possess the ability to diffuse readily in different materials and become uniformly distributed within a defined space. "Biologic gasses" are assumed to diffuse freely across biologic membranes, acting in a variety of functional capacities [70]; hydrogen gas is an example of this.

The gaseous diffusion of H_2 is indeed proven by monitoring its concentration inside various tissues. H_2 can be detected with specific electrodes. H_2 concentration has been monitored within a rat myocardium. The electrode was inserted into the non-ischemic myocardium of the left ventricle. The incremental rate of H_2 saturation for the non-ischemic myocardium and arterial blood was similar. Then, the electrode was inserted into the 'at risk' area for infarction to investigate the diffusion of H_2 into the ischemic myocardium, induced by coronary artery occlusion. Notably, H_2 concentration was increased even in the ischemic myocardium. Although the incremental rate of H_2 saturation was slower in the ischemic myocardium than in the non-ischemic myocardium, the peak level of H_2 in the ischemic myocardium was approximately two thirds of the value observed for the non-ischemic myocardium [58].

6.3. Protective Effects on Ischemia Reperfusion Model by Rat Cerebral Infarction

Hydrogen gas was applied to a rat model of ischemiareperfusion as an acute model [82]. We produced focal ischemia by occlusion of the rat middle cerebral artery with subsequent reperfusion. One day after middle cerebral artery occlusion, infarct volumes decreased in a H₂-dependent manner. One week after middle cerebral artery occlusion, the difference in infarct volumes between non-treated and H₂-treated rats increased. H₂-treated rats also showed improvements in body weight and temperature and movement defects vs. untreated rats. Thus, H₂ suppressed not only the initial brain injury, but also the progression of injury. H₂ markedly decreased several oxidative stress markers. In this experiment, H₂ was demonstrated to have the potential to markedly decrease oxidative stress and suppress brain injury [12].

6.4. Protective Effects on Hepatic and Cardiac Ischemia Reperfusion Injury

Next, inhalation of hydrogen gas was also applied to a hepatic ischemia reperfusion injury model [83]. Inhalation of H_2 clearly attenuated the degeneration induced by hepatic ischemia reperfusion and increased the protective effect in an H_2 -dependent manner. In contrast, helium gas (He) exhibited no effect, indicating that H_2 clearly has a specific protective effect [84].

The degree of cardioprotection against ischemia-reperfusion injury was evaluated by measuring oxidative damage and infarct size after left anterior descending coronary artery occlusion and reperfusion. Inhalation of an incombustible level of hydrogen gas (2%) before reperfusion significantly reduced oxidative stress-induced myocardial injury and infarct size without affecting hemodynamic parameters, and thereby prevented deleterious left ventricle remodeling [58].

6.5. Protective Effects in Organ Transplantation

H₂ inhalation significantly ameliorated intestinal and pulmonary transplant injury and prevented remote organ inflammation via its antioxidant effects [85, 86]. Ischemia/reperfusion injury during small intestinal and lung transplantation frequently causes complications, including dysmotility, inflammation and organ failure.

 $\rm H_2$ treatment resulted in significantly improved gastrointestinal transit, as well as jejunal smooth muscle contractility in response to bethanechol [86]. Graft lipid peroxidation was significantly reduced in the presence of $\rm H_2$, demonstrating antioxidant effects of $\rm H_2$ in the transplanted lungs. Exposure to 2% hydrogen gas significantly blocked the production of several pro-inflammatory mediators and reduced apoptosis with induction of the anti-apoptotic molecules B-cell lymphoma-2 and B-cell lymphoma-extra large.

Rat cardiac cold ischemia reperfusion injury was ameliorated with inhaled H_2 or carbon monoxide (CO), or both. Combined therapy with H_2 and CO demonstrated enhanced therapeutic efficacy via both anti-oxidant and anti-inflammatory mechanisms, and may be a clinically feasible approach for preventing cold ischemia reper-

fusion injury of the myocardium [87]. Inhaled hydrogen gas effectively reduced ventilator-induced lung injury-associated inflammatory responses, at both a local and systemic level, via its antioxidant, anti-inflammatory and anti-apoptotic effects [88].

6.6. Protective Effects in Infectious Diseases and antiinflammatory Effects

Sepsis, a multiple organ dysfunction syndrome, is the leading cause of death in critically ill patients [89]. Hydrogen gas inhalation significantly improved the survival rate and organ damage of septic mice with moderate or severe cecal ligation and puncture by reducing levels of early and late pro-inflammatory cytokines in serum and tissues [90].

The effects of 2% H_2 treatment was investigated on the survival rate and organ damage in zymosan-induced generalized inflammation model. The beneficial effects of H_2 treatment zymosan-induced organ damage were associated with decreased levels of oxidative product, increased activities of antioxidant enzyme, and reduced levels of early and late pro-inflammatory cytokines in serum and tissues. H_2 treatment protected against multiple organ damage in a zymosan-induced generalized inflammation model, suggesting the potential use of H_2 as a therapeutic agent in the therapy of conditions associated with inflammation-related multiple organ dysfunction syndrome [91].

6.7. Others

Other reports had the following titles: Hydrogen therapy reduces apoptosis in neonatal hypoxia-ischemia rat model [92]; hydrogen gas reduced acute hyperglycemia-enhanced hemorrhagic transformation in a focal ischemia rat model [93]; hydrogen is neuroprotective and preserves cerebrovascular reactivity in asphyxiated newborn pigs [94]; beneficial effects of hydrogen gas in a rat model of traumatic brain injury via reducing oxidative stress[95]; beneficial effects of hydrogen gas against spinal cord ischemia-reperfusion injury in rabbits [96]; and hydrogen protects vestibular hair cells from free radicals [97].

7. METHODS OF INGEST HYDROGEN II: ORAL INGESTION OF HYDROGEN WATER

7.1. Oral Ingestion by Drinking Hydrogen Water

Since inhaled hydrogen gas acts more rapidly, it may be suitable for defense against acute oxidative stress. In particular, inhalation of gas does not affect blood pressure; blood pressure increased by infusion may be serious in myocardial infarction; however, inhalation of hydrogen gas may be unsuitable or not practical as continuous H₂ consumption in daily life for preventive use. In contrast, solubilized H₂ (H₂-dissolved water; namely, hydrogen water) may be beneficial since it is a portable, easily administered and a safe means of delivering H₂ [98]. H₂ can be dissolved in water up to 0.8 mM under atmospheric pressure at room temperature as mentioned earlier. Unexpectedly, drinking hydrogen water had effects comparable to hydrogen gas inhalation [99].

Hydrogen water can be made by several methods, including dissolving hydrogen gas in water under high pressure, dissolving electrolyzed H_2 in water, and by the reaction of magnesium metal with water. The method of dissolving hydrogen gas under high pressure has an advantage because it is applicable not only using water but also any other solvents.

When water saturated with H_2 was placed into the stomach of a rat, H_2 was detected at several μM level in blood [98, 99]. Moreover, hepatic H_2 was monitored with a needle-type hydrogen electrode, and H_2 accumulated after oral administration of hydrogen water, partly explaining why consumption of even a small amount of H_2 over a short dwell time could efficiently improve various disease models. An additional *in vitro* experiment confirmed that

polymers of carbohydrates, including glycogen and starch, have an affinity for H₂ [99].

7.2. Prevention of Cognitive Decline

Chronic physical restraint stress on mice enhanced levels of oxidative stress in the brain, and impaired learning and memory [100, 101]. Consumption of hydrogen water ad libitum suppressed the increase in oxidative stress, and prevented cognitive impairment. Neural proliferation in the dentate gyrus of the hippocampus was suppressed by restraint stress [101]. The consumption of hydrogen water ameliorated the reduced proliferation; however, a mechanistic link between H2-dependent changes in neurogenesis and cognitive impairments remains unclear. Thus, continuous consumption of hydrogen water reduced oxidative stress in the brain and prevented the stress-induced decline in learning and memory [98].

7.3. Preventive and Therapeutic Affects on Parkinson Disease Model

In Parkinson's disease, mitochondrial dysfunction and the associated oxidative stress are major causes of dopaminergic cell loss in the substantia nigra [102]. H₂ in drinking water was given before or after stereotactic surgery for 6-hydroxydopamine-induced nigrostrital degeneration in a rat model of Parkinson's disease. Hydrogen water prevented both the development and progression of nigrostriatal degeneration. Hydrogen water likely retards the development and progression of Parkinson's disease [103].

Drinking hydrogen water suppressed dopaminergic neuronal loss in another Parkinson's disease model induced by MPTP (1methyl-4-phenyl-1,2,3,6-tetrahydropyridine) [104].

7.4. Prevention of Atherosclerosis Model

Oxidative stress is involved in atherosclerosis [105, 106]; however most clinical trials of dietary antioxidants failed to show marked success in preventing atherosclerotic diseases [8, 107, 108]. Drinking hydrogen water ad libitum decreased the aorta oxidative stress level and prevented arteriosclerosis in an apolipoprotein E knockout mouse [109]. Thus, consumption of hydrogen water has potential to prevent arteriosclerosis more effectively than other antioxidants [110].

7.5. Improvement of Metabolic Syndrome

Increased oxidative stress in obesity affects metabolic syndrome [111]. Long-term drinking of hydrogen water significantly controlled fat and body weights, despite no change in the consumption of food and water. Moreover, drinking hydrogen water decreased levels of plasma glucose, insulin and triglyceride, the effect of which on hyperglycemia was similar to diet restriction [112]. A mechanistic study revealed that the gene expression of the hepatic hormone, fibroblast growth factor 21 (FGF21) was enhanced, which should function to enhance fatty acid and glucose expenditure. Indeed, drinking hydrogen water stimulated energy metabolism, as measured by O2 consumption and CO2 expiration. These results suggest the potential benefit of H₂ in improving obesity, diabetes and metabolic syndrome [112].

7.6. Prevention of Adverse Effects by an Anti-tumor Drug

Cisplatin is a widely used anti-cancer drug in the treatment of a wide range of tumors; however, its application is limited by causing nephrotoxicity, which may be mediated by oxidative stress [113]. Inhalation of hydrogen gas (1% H₂ in air) or drinking hydrogen water improved mortality and body-weight loss caused by cisplatin, and alleviated nephrotoxicity. Consumption of hydrogen water improved metamorphosis accompanying decreased apoptosis in the kidney. Despite its protective effects against cisplatin-induced toxicity, H2 did not impair the anti-tumor activity of cisplatin against cancer cell lines in vitro and in tumor-bearing mice in vivo. Thus, H₂, whether hydrogen gas or hydrogen water, could improve the quality of life of patients during chemotherapy [99]. This finding was confirmed by another group [114].

7.7. Anti-allergic Reactions

It was demonstrated using a mouse model that drinking hydrogen water could attenuate an immediate-type allergic reaction by suppressing the phosphorylation of FceRI-associated Lyn and its downstream signaling molecules, which subsequently inhibited NADPH oxidase activity and reduced the generation of hydrogen peroxide [115]. These findings imply that the beneficial effects of H₂ are not only imparted by its radical scavenging activity, but also by modulating a specific signaling pathway.

7.8. Effects on Transplantation

ROS contributes to the development of interstitial fibrosis and tubular atrophy seen in chronic allograft nephropathy. Nakao's group tested the effect of treatment with hydrogen water in a model of kidney transplantation, in which allografts from Lewis rats were orthotopically transplanted into Brown Norway recipients that had undergone bilateral nephrectomy. Drinking hydrogen water improved allograft function, slowed the progression of chronic allograft nephropathy, reduced oxidant injury and inflammatory mediator production, and improved overall survival. Inflammatory signaling pathways, such as mitogen-activated protein kinases, were less activated in renal allografts from hydrogen water-treated rats as compared with normal water-treated rats. Thus, oral hydrogen water is an effective antioxidant and anti-inflammatory agent that reduced chronic allograft nephropathy, improving the survival of rat renal allografts [116].

7.9. Others

It has been shown that drinking hydrogen water prevents superoxide formation in brain slices of vitamin C-depleted SMP30/GNL knockout mice [117], that H₂ in drinking water attenuates noiseinduced hearing loss in guinea pigs [118], that drinking hydrogen water ameliorated cognitive impairment in senescence-accelerated mice [119], and that H₂ exhibited potential cardioprotective effects in irradiated mice [120].

8. METHODS OF INGEST HYDROGEN III: INJECTION OF HYDROGEN SALINE

8.1. Advantage of injection

Even though oral administration is safe and convenient, H₂ in water tends to escape over time and some H2 is lost in the stomach or intestine, making it difficult to control the concentration of H₂ administered. Administration of H₂ via an injectable hydrogen saline (H₂-dissolved saline) vehicle may allow the delivery of more accurate concentrations of H₂ [121].

8.2. Effects of Hydrogen Saline on Various Disease Models

Sun's group administered H₂-saturated saline by peritoneal injection to various model animals with great success. Thus, hydrogen saline has potential in actual clinical treatment. For example, injection of hydrogen saline showed neuroprotective effects in a neonatal hypoxia-ischemia rat model [121]. Moreover, H₂ saline was applied to an Alzheimer's disease model mouse, which was generated by intracerebroventricular injection of the A\beta 1-42 peptide. H2 treatment decreased the level of oxidative stress and inflammation markers and prevented memory dysfunction and motor dysfunction, respectively [122].

They and other groups have demonstrated effects on many disease models, as published in the following reports [123-130].

9. METHODS OF INGEST HYDROGEN IV: DIRECT ABSORPTION OF HYDROGEN

9.1. Improvement of Glaucoma Model

Alternatively, H_2 -loaded eye drops were prepared by dissolving H_2 in saline and directly administering to the ocular surface [131, 132].

In acute glaucoma of the eyes, transient elevation of intraocular pressure causes significant reductions in the thickness of the retina by ischemia-reperfusion injury mediated through the generation of reactive oxygen species [133]. The direct application of eye drops containing H₂ ameliorated ischemia-reperfusion injury of the retina in a rat model. When H₂ eye drops were continuously administered, the H₂ concentration increased in the vitreous body and the •OH level decreased during retinal ischemia-reperfusion. H₂ eye drops reduced the number of apoptotic and oxidative stress marker-positive cells 1 day after ischemia-reperfusion injury, and reduced retinal thinning with accompanying activation of Müller glia, astrocytes and microglia at 7 days after ischemia-reperfusion injury, improving the recovery of inner retinal layer thickness to >70%.

Moreover, we devised eye drops with dissolved H_2 to directly administer H_2 to the retina, and monitored the time course of changes in H_2 levels using a needle-shaped hydrogen sensor electrode inserted through the sclera to the vitreous body in rats. H_2 was able to reach the vitreous body by administering H_2 saturated in normal saline. When H_2 eye drops were administered continuously, approximately 70% H_2 was detected on the ocular surface. Two minutes after the start of administration, H_2 concentration in the vitreous body started to increase and reached a maximum level after 15 min. At that time, H_2 concentration was approximately 20% of H_2 in the eye-drops. The maximum concentration of H_2 in the vitreous body reached approximately one third of the value observed on the ocular surface [131].

9.2. Hydrogen Bath

 H_2 easily penetrates the skin and distributes throughout the body via blood flow. Thus, taking a warm water bath with dissolved H_2 is a method of incorporating H_2 into the body in daily life, especially in Japan. It takes only 10 minutes to distribute throughout the whole body, as judged by measuring hydrogen gas in expiration (unpublished results).

10. METHODS OF INGEST HYDROGEN V: INCREASE IN INTESTINAL HYDROGEN

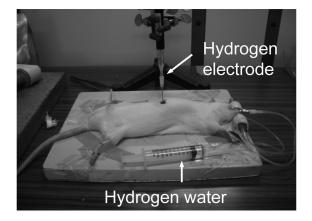
10.1. Production of Hydrogen in Intestinal Bacteria

Other medical gasses, CO, NO and H₂S, are generated by endogenous enzymatic systems. Pharmaceutical development has taken advantage of these systems to design exogenous molecules to simulate those generated endogenously; however, mammals lack their own enzyme to produce H₂ [70].

Instead of endogenous enzymatic systems, the spontaneous production of hydrogen gas in the human body occurs via the fermentation of undigested carbohydrates by resident enterobacterial flora [134]. H_2 is transferred to the portal circulation and excreted through the breath in significant amounts [135]. For this reason, measurement of H_2 levels in expired air is used to detect carbohydrate malabsorption [76]; however, there have been few studies on the physiological function of gastrointestinal tract-derived hydrogen gas as an antioxidant.

10.2. Are α-glucosidase Inhibitors an Indirect Antioxidant?

 α -Glucosidase inhibitors are pharmacological agents that specifically reduce postprandial hyperglycemia through retardation of disaccharide digestion, thereby reducing glucose absorption. A large scale epidemiologic trial has demonstrated that the treatment of patients with impaired glucose tolerance with an α -glucosidase



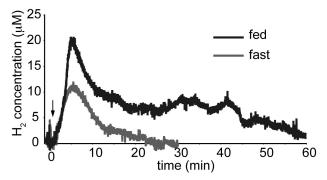


Fig. (2). Measurement of the accumulation of H_2 in rat liver.

The concentration of H_2 in the liver was monitored using a needle-type hydrogen sensor inserted into fed- or overnight fasted-rat liver. Rat received hydrogen water (0.8 mM H_2 in water) orally by stomach gavage at 15 ml/kg. Arrow indicates the time point when rat was administered hydrogen water.

inhibitor was associated with a 25% reduction in the risk of progression to diabetes, a 34% reduction in the risk of developing de novo hypertension, and a 49% risk reduction of cardiovascular events [136]. Furthermore, meta-analysis of seven long-term studies suggested that acarbose reduced the risk of myocardial infarction for patients with type 2 diabetes [137]. Such risk reduction for coronary heart disease events in patients with type 2 diabetes was not observed by improved glycemic control achieved by intensified treatment with insulin and glibenclamid [138]. Actually, acarbose, which is an α -glucosidase inhibitor, markedly increased H_2 production in volunteers. Thus, we propose that H_2 produced by intestinal bacteria acts as a unique antioxidant and prevents cardiovascular events [139].

10.3. Anti-inflammation Effects by Intestinal Bacteria via Hydrogen

Escherichia coli can produce a considerable amount of H_2 by catalyzing with hydrogenase. Kawai et al. examined whether H_2 released from intestinally colonized bacteria could affect concanavalin A-induced mouse hepatitis. Reconstitution of intestinal flora with H_2 -producing E. coli, but not hydrogenase-deficient mutant E. coli, down-regulated concanavalin A-induced liver inflammation. These results indicate that H_2 released from intestinal bacteria can suppress inflammation [140]. H_2 also mediates the suppression of colon inflammation induced by dextran sodium sulfate [141].

10.4. Others

Dietary turmeric induced H₂ production from the intestinal bacteria [142], and lactulose was shown to be an indirect antioxidant ameliorating inflammatory bowel disease [143].

11. CLINICAL TESTS

Several groups have started clinical examinations. Clinical tests have revealed that drinking hydrogen water reduced oxidative stress markers in patients with type 2 diabetes [144] or subjects with potential metabolic syndrome [145] and influenced glucose [144] and cholesterol metabolism [145].

Hemodialysis using dialysis solution with H_2 significantly decreased the levels of plasma monocyte chemoattractant protein 1 and myeloperoxidase [146].

12. REGULATION OF GENE EXPRESSIONS AND PROTEIN PHOSPHORYLATIONS

It has been reported that H_2 acts as an anti-inflammatory and anti-allergic regulator by inducing inflammatory cytokines and inhibiting phosphorylating signal factors, respectively; however, the transcriptional factors and kinases involved in the effects afforded by H_c have not been identified.

 H_2 decreased the expressions of pro-inflammatory factors, including TNF-α, IL-6, IL-1β, CCL2 and IL-10, TNF-γ, IL-12, ICAM-1 [85], HMGB-1 [147], NF-κB [148], PGE2, and PGE2 [54].

Moreover, H₂ up- or down-regulated the factors involved in apoptosis toward the inhibition of apoptosis: H₂ suppressed the

expressions of pro-apoptotic factors, including casapase 3 [92, 149], and caspase 12 [92], caspase 8 [86] and BAX [86]. Conversely $\rm H_2$ stimulated the expressions of the anti-apoptoptic factors of Bcl-2 and Bcl-xL [86].

 H_2 is involved in the regulation of various factors; up-regulation of PCNA, bFGF, HGF, IFN- γ , and down-regulation of i-NOS [87] and VEGF [54].

As a signal transduction contributor, H_2 inhibited the phospahorylations of some signal proteins, including MEK, p38, ERK, JNK [116] and Lyn, Syk, PLC γ 1, γ 2, Akt, ERK1/2, JNK, p38, cPLA2, ASK1, I κ B α [115].

Heme oxygenase-1 (HO-1), a microsomal enzyme degrading heme to carbon monoxide, free iron, and biliverdin, participates in the cell defense against oxidative stress and has been speculated to be a new therapeutic target [150]. Notably, H₂ modulates HO-1 expression, which is commonly up-regulated by these medical gasses [48, 151]. Additionally, H₂ up-reguated the expression of FGF21, which is a regulator of energy metabolism [112].

As essential questions, it remains unknown how H_2 regulates gene expressions and phosphorylations, and whether the above regulations of transcription and phosphorylation are the cause or consequence of the effects of H_2 . The primary molecular target of H_2 remains unknown.

Table 1. Diseases and Physiological States for Which Hydrogen Effects are Reported as Classified by Target Organs [152].

| Disease/Physiology | Species | Source of H ₂ | Reference |
|---------------------------------|---------|--------------------------|------------|
| Brain | | | |
| Cerebral infarction | rodent | gas | [12] |
| Superoxide in brain | rodent | water | [117] |
| Neonatal brain hypoxia | rodent | gas | [92] |
| | rodent | saline | [131] |
| | pig | gas | [94] |
| Restraint-induced dementia | rodent | water | [98] |
| Alzheimer's disease | rodent | saline | [122] |
| Senile dementia | rodent | water | [119] |
| Parkinson's disease | rodent | water | [103, 104] |
| Hemorrhagic cerebral infarction | rodent | gas | [93] |
| Traumatic brain injury | rodent | gas | [95] |
| Spinal cord | | | |
| Spinal cord injury | rodent | saline | [130] |
| Eye | | | |
| Glaucoma | rodent | eye drops | [131] |
| Corneal alkali burn | rodent | eye drops | [132] |
| Ear | | | |
| Hearing disturbance | rodent | medium | [50] |
| | rodent | gas | [97] |
| | rodent | water | [118] |

(Table 1) Contd....

| Disease/Physiology | Species | Source of H ₂ | Reference |
|----------------------------------|---------------|--------------------------|------------|
| Lung | | | |
| Oxygen-induced lung injury | rodent | saline | [128, 129] |
| Lung transplantation | rodent | gas | [86] |
| Heart | | | |
| Myocardial infarction | rodent | gas | [58] |
| | rodent | saline | [149] |
| Heart transplantation | rodent | gas | [87] |
| Irradiation-induced heart injury | rodent | water | [120] |
| Liver | | | |
| Hepatic ischemia | rodent | gas | [84] |
| Hepatitis | rodent | bacteria | [140] |
| Obstructive jaundice | rodent | saline | [124] |
| Kidney | | | |
| Cisplatin nephropathy | rodent | gas, water | [99] |
| | rodent | water | [114] |
| Hemodialysis | human | dialysis | [146] |
| Kidney transplantation | rodent | water | [116] |
| Pancreas | | | |
| Acute pancreatitis | rodent | saline | [148] |
| Intestine | | | |
| Intestinal graft | rodent | gas | [85] |
| | rodent | saline | [125, 130] |
| Ulcerative colitis | rodent | gas | [141] |
| Blood vessel | | | |
| Atherosclerosis | rodent | water | [110] |
| Metabolism | | | |
| Diabetes mellitus type 2 | human | water | [144] |
| Metabolic syndrome | human | water | [145] |
| Obesity/Diabetes | rodent | water | [112] |
| Inflammation and allergy | | | |
| Allergy type I | rodent | water | [115] |
| Sepsis | rodent | gas | [90] |
| Zymosan-induced inflammation | rodent | gas | [91] |
| Others | | | |
| Multipotent stromal cells | cells | medium | [54] |
| Radiation injury | cells, rodent | medium | [52] |

13. CLOSING REMARKS: ISSUES TO BE DISSOLVED IN THE FUTURE

In our first report published in 2007, we indicated that H_2 reacted with strong reactive oxygen/nitrogen species, including \bullet OH and ONOO $^-$ in cell-free reactions. Cells cultured in H_2 -rich medium were protected against oxidative stress by the \bullet OH-scavenging activity of H_2 , depending upon the decrease of \bullet OH [12]; however, recent evidence shows that the scavenging property is not the only explanation for the potent beneficial effects of H_2 . When model animals and human subjects consumed H_2 by drinking water with dissolved H_2 , even a very small amount of H_2 was extensively effective. It may be difficult to explain that direct reduction of \bullet OH by a very small amount of H_2 reveals all the functions of H_2 , because the saturated level of H_2 is only 0.8 mM and the dwelling time of \bullet OH is very short in the body.

We have recently shown that H_2 can be accumulated with hepatic glycogen; this finding indicates the possible accumulation of H_2 in a specific region; however, it is unlikely that the amount of H_2 is sufficient to exhibit all of its functions [112]. Additionally, drinking 0.04 or 0.08 mM H_2 was shown to be effective [104, 112]. The amount of administered H_2 seems to be, in many cases, independent of the magnitude of effects. Intestinal bacteria produce more than 1 liter of hydrogen gas per day, whereas the amount of H_2 originating from drinking hydrogen water is less than 50 ml. Nevertheless, additional H_2 in drinking hydrogen water is unambiguously effective.

Many additional issues of hydrogen therapy including the molecular mechanisms underlying the marked effects of a very small amount of H_2 remain elusive. The primary molecular target of H_2 remains unknown. Although H_2 regulates various gene expressions and protein-phosphorylations, it remains unclear whether such regulations are the cause or consequence of the effects against oxidative stress. One of the open questions is how H_2 involves the cross-talk among anti-oxidation, anti-inflammation and anti-allergy. Thus, it should not be fair to classify the roles of H_2 by outward effects at this stage.

Finally, the author summarizes the reports showing the effects of H_2 by the classification of target organs (Table 1) [152].

DISCLOSURE

The author declares no conflicts of interest.

REFERENCES

- Wallace DC. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. Annu Rev Genet 2005; 39: 359-407.
- [2] Ferrari R, Ceconi C, Curello S, Cargnoni A, Pasini E, Visioli O. The occurrence of oxidative stress during reperfusion in experimental animals and men. Cardiovasc Drugs Ther 1991; 5 Suppl 2: 277-87.
- [3] Andersen JK. Oxidative stress in neurodegeneration: cause or consequence? Nat Med 2004; 10 Suppl: S18-25.
- [4] Ohta S, Ohsawa I. Dysfunction of mitochondria and oxidative stress in the pathogenesis of Alzheimer's disease: on defects in the cytochrome c oxidase complex and aldehyde detoxification. J Alzheimers Dis 2006: 9: 155-66.
- [5] Ohta S, Ohsawa I, Kamino K, Ando F, Shimokata H. Mitochondrial ALDH2 deficiency as an oxidative stress. Ann N Y Acad Sci 2004; 1011: 36-44.
- [6] Chang JC, Kou SJ, Lin WT, Liu CS. Regulatory role of mitochondria in oxidative stress and atherosclerosis. World J Cardiol 2010; 2: 150-9.
- [7] Finkel T, Holbrook NJ. Oxidants, oxidative stress and the biology of ageing. Nature 2000; 408: 239-47.
- [8] Steinhubl SR. Why have antioxidants failed in clinical trials? Am J Cardiol 2008; 101: 14D-9D.
- [9] Hercberg S, Kesse-Guyot E, Druesne-Pecollo N, et al. Incidence of cancers, ischemic cardiovascular diseases and mortality during 5year follow-up after stopping antioxidant vitamins and minerals

- supplements: a postintervention follow-up in the SU.VI.MAX Study. Int J Cancer 2010; 127: 1875-81.
- [10] Brambilla D, Mancuso C, Scuderi MR, et al. The role of antioxidant supplement in immune system, neoplastic, and neurodegenerative disorders: a point of view for an assessment of the risk/benefit profile. Nutr J 2008: 7: 29.
- [11] Hackam DG. Review: antioxidant supplements for primary and secondary prevention do not decrease mortality. ACP J Club 2007; 147: 4.
- [12] Ohsawa I, Ishikawa M, Takahashi K, et al. Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. Nat Med 2007; 13: 688-94.
- [13] Salganik RI. The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients and the human population. J. Am. Coll. Nutr 2001; 20: 464S-72S; discussion 73S-75S.
- [14] Sauer H, Wartenberg M, Hescheler J. Reactive oxygen species as intracellular messengers during cell growth and differentiation. Cell. Physiol. Biochem 2001; 11: 173-86.
- [15] Liu H, Colavitti R, Rovira, II, Finkel T. Redox-dependent transcriptional regulation. Circ. Res 2005; 97: 967-74.
- [16] Harma MI, Harma M, Erel O. Measuring plasma oxidative stress biomarkers in sport medicine. Eur J Appl Physiol 2006; 97: 505; author reply 6-8.
- [17] Tanriverdi H, Evrengul H, Kuru O, et al. Cigarette smoking induced oxidative stress may impair endothelial function and coronary blood flow in angiographically normal coronary arteries. Circ J 2006; 70: 593-9.
- [18] Grassi D, Desideri G, Ferri L, Aggio A, Tiberti S, Ferri C. Oxidative stress and endothelial dysfunction: say no to cigarette smoking! Curr Pharm Des 2010; 16: 2539-50.
- [19] Agarwal R. Smoking, oxidative stress and inflammation: impact on resting energy expenditure in diabetic nephropathy. BMC Nephrol 2005; 6: 13.
- [20] Turrens JF. Mitochondrial formation of reactive oxygen species. J Physiol 2003; 552: 335-44.
- [21] Lin MT, Beal MF. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. Nature 2006; 443: 787-95.
- [22] Halliwell B, Gutteridge JM. Biologically relevant metal iondependent hydroxyl radical generation. An update. FEBS Lett 1992: 307: 108-12.
- [23] Peled-Kamar M, Lotem J, Wirguin I, Weiner L, Hermalin A, Groner Y. Oxidative stress mediates impairment of muscle function in transgenic mice with elevated level of wild-type Cu/Zn superoxide dismutase. Proc Natl Acad Sci U S A 1997; 94: 3883-7.
- [24] Chan PH, Epstein CJ, Li Y, et al. Transgenic mice and knockout mutants in the study of oxidative stress in brain injury. J Neurotrauma 1995; 12: 815-24.
- [25] Mitsui A, Hamuro J, Nakamura H, et al. Overexpression of human thioredoxin in transgenic mice controls oxidative stress and life span. Antioxid Redox Signal 2002; 4: 693-6.
- [26] Stefanova N, Reindl M, Neumann M, et al. Oxidative stress in transgenic mice with oligodendroglial alpha-synuclein overexpression replicates the characteristic neuropathology of multiple system atrophy. Am J Pathol 2005; 166: 869-76.
- [27] Schriner SE, Linford NJ, Martin GM, et al. Extension of murine life span by overexpression of catalase targeted to mitochondria. Science 2005; 308: 1909-11.
- [28] Stockl P, Zankl C, Hutter E, et al. Partial uncoupling of oxidative phosphorylation induces premature senescence in human fibroblasts and yeast mother cells. Free Radic Biol Med 2007; 43: 947-58.
- [29] Jia JJ, Zhang X, Ge CR, Jois M. The polymorphisms of UCP2 and UCP3 genes associated with fat metabolism, obesity and diabetes. Obes Rev 2009; 10: 519-26.
- [30] Jia JJ, Tian YB, Cao ZH, et al. The polymorphisms of UCP1 genes associated with fat metabolism, obesity and diabetes. Mol. Biol. Rep 2010; 37: 1513-22.
- [31] Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ. Res 2010; 107: 1058-70.
- [32] Ohsawa I, Nishimaki K, Murakami Y, Suzuki Y, Ishikawa M, Ohta S. Age-dependent neurodegeneration accompanying memory loss in transgenic mice defective in mitochondrial aldehyde dehydrogenase 2 activity. J Neurosci 2008; 28: 6239-49.
- [33] Endo J, Sano M, Katayama T, Hishiki T, et al. Metabolic remodeling induced by mitochondrial aldehyde stress stimulates

- tolerance to oxidative stress in the heart. Circ Res 2009; 105: 1118-27
- [34] Reuter S, Gupta SC, Chaturvedi MM, Aggarwal BB. Oxidative stress, inflammation, and cancer: how are they linked? Free Radic Biol Med 2010; 49: 1603-16.
- [35] Vaziri ND, Rodriguez-Iturbe B. Mechanisms of disease: oxidative stress and inflammation in the pathogenesis of hypertension. Nat Clin Pract Nephrol 2006; 2: 582-93.
- [36] Bolli R, Jeroudi MO, Patel BS, et al. Marked reduction of free radical generation and contractile dysfunction by antioxidant therapy begun at the time of reperfusion. Evidence that myocardial "stunning" is a manifestation of reperfusion injury. Circ Res 1989; 65: 607-22.
- [37] Zweier JL. Measurement of superoxide-derived free radicals in the reperfused heart. Evidence for a free radical mechanism of reperfusion injury. J Biol Chem 1988; 263: 1353-7.
- [38] Bolli R, Patel BS, Jeroudi MO, Lai EK, McCay PB. Demonstration of free radical generation in "stunned" myocardium of intact dogs with the use of the spin trap alpha-phenyl N-tert-butyl nitrone. J Clin Invest 1988; 82: 476-85.
- [39] Vanden Hoek T, Becker LB, Shao ZH, Li CQ, Schumacker PT. Preconditioning in cardiomyocytes protects by attenuating oxidant stress at reperfusion. Circ Res 2000; 86: 541-8.
- [40] Halliwell B, Gutteridge JM. Oxygen free radicals and iron in relation to biology and medicine: some problems and concepts. Arch. Biochem. Biophys 1986; 246: 501-14.
- [41] Halestrap AP, Clarke SJ, Khaliulin I. The role of mitochondria in protection of the heart by preconditioning. Biochim Biophys Acta 2007; 1767: 1007-31.
- [42] Flaherty JT, Pitt B, Gruber JW, Heuser RR, Rothbaum DA, Burwell LR, George BS, Kereiakes DJ, Deitchman D, Gustafson N, et al. Recombinant human superoxide dismutase (h-SOD) fails to improve recovery of ventricular function in patients undergoing coronary angioplasty for acute myocardial infarction. Circulation 1994; 89: 1982-91.
- [43] Richard VJ, Murry CE, Jennings RB, Reimer KA. Therapy to reduce free radicals during early reperfusion does not limit the size of myocardial infarcts caused by 90 minutes of ischemia in dogs. Circulation 1988; 78: 473-80.
- [44] Ristow M, Zarse K. How increased oxidative stress promotes longevity and metabolic health: The concept of mitochondrial hormesis (mitohormesis). Exp Gerontol 2010; 45: 410-8.
- [45] Ristow M, Zarse K, Oberbach A, et al. Antioxidants prevent health-promoting effects of physical exercise in humans. Proc Natl Acad Sci U S A 2009; 106: 8665-70.
- [46] Penna C, Rastaldo R, Mancardi D, et al. Post-conditioning induced cardioprotection requires signaling through a redox-sensitive mechanism, mitochondrial ATP-sensitive K+ channel and protein kinase C activation. Basic Res Cardiol 2006; 101: 180-9.
- [47] Downey JM, Cohen MV. A really radical observation--a comment on Penna et al. in Basic Res Cardiol (2006) 101:180-189. Basic Res Cardiol 2006; 101: 190-1.
- [48] Huang CS, Kawamura T, Toyoda Y, Nakao A. Recent advances in hydrogen research as a therapeutic medical gas. Free Radic Res 2010; 44: 971-82.
- [49] Setsukinai K, Urano Y, Kakinuma K, Majima HJ, Nagano T. Development of novel fluorescence probes that can reliably detect reactive oxygen species and distinguish specific species. J Biol Chem 2003; 278: 3170-5.
- [50] Kikkawa YS, Nakagawa T, Horie RT, Ito J. Hydrogen protects auditory hair cells from free radicals. Neuroreport 2009; 20: 689-94
- [51] Taura A, Kikkawa YS, Nakagawa T, Ito J. Hydrogen protects vestibular hair cells from free radicals. Acta Otolaryngol Suppl 2010: 95-100.
- [52] Qian L, Cao F, Cui J, *et al.* Radioprotective effect of hydrogen in cultured cells and mice. Free Radic Res 2010; 44: 275-82.
- [53] Schoenfeld MP, Ansari RR, Zakrajsek JF, et al. Hydrogen therapy may reduce the risks related to radiation-induced oxidative stress in space flight. Med Hypotheses 2011; 76: 117-8.
- [54] Kawasaki H, Guan J, Tamama K. Hydrogen gas treatment prolongs replicative lifespan of bone marrow multipotential stromal cells in vitro while preserving differentiation and paracrine potentials. Biochem Biophys Res Commun 2010; 397: 608-13.
- [55] Murphy MP, Smith RA. Drug delivery to mitochondria: the key to mitochondrial medicine. Adv Drug Deliv Rev 2000; 41: 235-50.

- [56] Murphy MP. Selective targeting of bioactive compounds to mitochondria. Trends Biotechnol 1997; 15: 326-30.
- [57] Smith RA, Murphy MP. Mitochondria-targeted antioxidants as therapies. Discov Med 2011; 11: 106-14.
- [58] Hayashida K, Sano M, Ohsawa I, et al. Inhalation of hydrogen gas reduces infarct size in the rat model of myocardial ischemiareperfusion injury. Biochem Biophys Res Commun 2008; 373: 30-5
- [59] Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG, Gluud C. Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and metaanalysis. JAMA 2007; 297: 842-57.
- [60] Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG, Gluud C. Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases. Cochrane Database Syst Rev 2008: CD007176.
- [61] Gray SL, Anderson ML, Crane PK, et al. Antioxidant vitamin supplement use and risk of dementia or Alzheimer's disease in older adults. J Am Geriatr Soc 2008; 56: 291-5.
- [62] Walker C. Antioxidant supplements do not improve mortality and may cause harm. Am Fam Physician 2008; 78: 1079-80.
- [63] Bjelakovic G, Gluud C. Surviving antioxidant supplements. J. Natl. Cancer Inst 2007; 99: 742-3.
- [64] Miller ER, 3rd, Pastor-Barriuso R, Dalal D, Riemersma RA, Appel LJ, Guallar E. Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. Ann. Intern. Med 2005; 142: 37-46.
- [65] Mandal CC, Ganapathy S, Gorin Y, et al. Reactive oxygen species derived from Nox4 mediate BMP2 gene transcription and osteoblast differentiation. Biochem J 2010; 433: 393-402.
- [66] Chandel NS, Maltepe E, Goldwasser E, Mathieu CE, Simon MC, Schumacker PT. Mitochondrial reactive oxygen species trigger hypoxia-induced transcription. Proc Natl Acad Sci U S A 1998; 95: 11715-20.
- [67] Carriere A, Carmona MC, Fernandez Y, et al. Mitochondrial reactive oxygen species control the transcription factor CHOP-10/GADD153 and adipocyte differentiation: a mechanism for hypoxia-dependent effect. J Biol Chem 2004; 279: 40462-9.
- [68] Winterbourn CC. Biological reactivity and biomarkers of the neutrophil oxidant, hypochlorous acid. Toxicology 2002; 181-182: 223-7.
- [69] Murad F. Discovery of some of the biological effects of nitric oxide and its role in cell signaling. Biosci. Rep 2004; 24: 452-74.
- [70] Kajimura M, Fukuda R, Bateman RM, Yamamoto T, Suematsu M. Interactions of multiple gas-transducing systems: hallmarks and uncertainties of CO, NO, and H2S gas biology. Antioxid Redox Signal 2010; 13: 157-92.
- [71] Motterlini R, Otterbein LE. The therapeutic potential of carbon monoxide. Nat Rev Drug Discov 2010; 9: 728-43.
- [72] Kimura H. Hydrogen sulfide: from brain to gut. Antioxid Redox Signal 2010; 12: 1111-23.
- [73] Szabo C. Hydrogen sulphide and its therapeutic potential. Nat Rev Drug Discov 2007; 6: 917-35.
- [74] Elrod JW, Calvert JW, Morrison J, et al. Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function. Proc Natl Acad Sci U S A 2007; 104: 15560-5
- [75] Foresti R, Bani-Hani MG, Motterlini R. Use of carbon monoxide as a therapeutic agent: promises and challenges. Intensive Care Med
- [76] Kobayashi A, Ishikawa K, Matsumoto H, Kimura S, Kamiyama Y, Maruyama Y. Synergetic antioxidant and vasodilatory action of carbon monoxide in angiotensin II - induced cardiac hypertrophy. Hypertension 2007; 50: 1040-8.
- [77] Bloch KD, Ichinose F, Roberts JD, Jr., Zapol WM. Inhaled NO as a therapeutic agent. Cardiovasc Res 2007; 75: 339-48.
- [78] Abraini JH, Gardette-Chauffour MC, Martinez E, Rostain JC, Lemaire C. Psychophysiological reactions in humans during an open sea dive to 500 m with a hydrogen-helium-oxygen mixture. J Appl Physiol 1994; 76: 1113-8.
- [79] Lillo RS, Parker EC, Porter WR. Decompression comparison of helium and hydrogen in rats. J Appl Physiol 1997; 82: 892-901.
- [80] Lillo RS, Parker EC. Mixed-gas model for predicting decompression sickness in rats. J Appl Physiol 2000; 89: 2107-16.
- [81] Fontanari P, Badier M, Guillot C, et al. Changes in maximal performance of inspiratory and skeletal muscles during and after

- the 7.1-MPa Hydra 10 record human dive. Eur J Appl Physiol 2000; 81: 325-8.
- [82] Peters O, Back T, Lindauer U, et al. Increased formation of reactive oxygen species after permanent and reversible middle cerebral artery occlusion in the rat. J Cereb Blood Flow Metab 1998; 18: 196-205.
- [83] Jaeschke H, Smith CV, Mitchell JR. Reactive oxygen species during ischemia-reflow injury in isolated perfused rat liver. J. Clin. Invest 1988; 81: 1240-6.
- [84] Fukuda K, Asoh S, Ishikawa M, Yamamoto Y, Ohsawa I, Ohta S. Inhalation of hydrogen gas suppresses hepatic injury caused by ischemia/reperfusion through reducing oxidative stress. Biochem Biophys Res Commun 2007; 361: 670-4.
- [85] Buchholz BM, Kaczorowski DJ, Sugimoto R, et al. Hydrogen inhalation ameliorates oxidative stress in transplantation induced intestinal graft injury. Am J Transplant 2008; 8: 2015-24.
- [86] Kawamura T, Huang CS, Tochigi N, et al. Inhaled hydrogen gas therapy for prevention of lung transplant-induced ischemia/reperfusion injury in rats. Transplantation 2010; 90: 1344-51.
- [87] Nakao A, Kaczorowski DJ, Wang Y, et al. Amelioration of rat cardiac cold ischemia/reperfusion injury with inhaled hydrogen or carbon monoxide, or both. J Heart Lung Transplant 2010; 29: 544-53
- [88] Huang CS, Kawamura T, Lee S, et al. Hydrogen inhalation ameliorates ventilator-induced lung injury. Crit Care 2010; 14: R234.
- [89] Victor VM, Espulgues JV, Hernandez-Mijares A, Rocha M. Oxidative stress and mitochondrial dysfunction in sepsis: a potential therapy with mitochondria-targeted antioxidants. Infect Disord Drug Targets 2009; 9: 376-89.
- [90] Xie KL, Yu YH, Pei YP, et al. Protective effects of hydrogen gas on murine polymicrobial sepsis via reducing oxidative stress and HMGB1 release. Shock 2010; 34: 90-7.
- [91] Xie K, Yu Y, Zhang Z, et al. Hydrogen gas improves survival rate and organ damage in zymosan-induced generalized inflammation model. Shock 2010; 34: 495-501.
- [92] Cai J, Kang Z, Liu WW, et al. Hydrogen therapy reduces apoptosis in neonatal hypoxia-ischemia rat model. Neurosci Lett 2008; 441: 167-72.
- [93] Chen CH, Manaenko A, Zhan Y, Liu WW, Ostrowki RP, Tang J, Zhang JH. Hydrogen gas reduced acute hyperglycemia-enhanced hemorrhagic transformation in a focal ischemia rat model. Neuroscience 2010; 169: 402-14.
- [94] Domoki F, Olah O, Zimmermann A, et al. Hydrogen is neuroprotective and preserves cerebrovascular reactivity in asphyxiated newborn pigs. Pediatr Res 2010; 68: 387-92.
- [95] Ji X, Liu W, Xie K, et al. Beneficial effects of hydrogen gas in a rat model of traumatic brain injury via reducing oxidative stress. Brain Res 2010; 1354: 196-205.
- [96] Huang Y, Xie K, Li J, et al. Beneficial effects of hydrogen gas against spinal cord ischemia-reperfusion injury in rabbits. Brain Res 2011; 1378: 125-36.
- [97] Taura A, Kikkawa YS, Nakagawa T, Ito J. Hydrogen protects vestibular hair cells from free radicals. Acta Otolaryngol. (Stockh) 2010; 130: 95-100.
- [98] Nagata K, Nakashima-Kamimura N, Mikami T, Ohsawa I, Ohta S. Consumption of molecular hydrogen prevents the stress-induced impairments in hippocampus-dependent learning tasks during chronic physical restraint in mice. Neuropsychopharmacology 2009; 34: 501-8.
- [99] Nakashima-Kamimura N, Mori T, Ohsawa I, Asoh S, Ohta S. Molecular hydrogen alleviates nephrotoxicity induced by an anticancer drug cisplatin without compromising anti-tumor activity in mice. Cancer Chemother Pharmacol 2009; 64: 753-61.
- [100] Liu J, Wang X, Shigenaga MK, Yeo HC, Mori A, Ames BN. Immobilization stress causes oxidative damage to lipid, protein, and DNA in the brain of rats. FASEB J 1996; 10: 1532-8.
- [101] Abrous DN, Koehl M, Le Moal M. Adult neurogenesis: from precursors to network and physiology. Physiol Rev 2005; 85: 523-69
- [102] Schapira AH. Mitochondria in the aetiology and pathogenesis of Parkinson's disease. Lancet Neurol 2008; 7: 97-109.
- [103] Fu Y, Ito M, Fujita Y, et al. Molecular hydrogen is protective against 6-hydroxydopamine-induced nigrostriatal degeneration in a rat model of Parkinson's disease. Neurosci Lett 2009; 453: 81-5.

- [104] Fujita K, Seike T, Yutsudo N, et al. Hydrogen in drinking water reduces dopaminergic neuronal loss in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. PLoS One 2009; 4: e7247.
- [105] Victor VM, Apostolova N, Herance R, Hernandez-Mijares A, Rocha M. Oxidative stress and mitochondrial dysfunction in atherosclerosis: mitochondria-targeted antioxidants as potential therapy. Curr Med Chem 2009; 16: 4654-67.
- [106] Stocker R, Keaney JF, Jr. Role of oxidative modifications in atherosclerosis. Physiol. Rev 2004; 84: 1381-478.
- [107] Upston JM, Kritharides L, Stocker R. The role of vitamin E in atherosclerosis. Prog. Lipid Res 2003; 42: 405-22.
- [108] Hodis HN, Mack WJ, LaBree L, et al. Alpha-tocopherol supplementation in healthy individuals reduces low-density lipoprotein oxidation but not atherosclerosis: the Vitamin E Atherosclerosis Prevention Study (VEAPS). Circulation 2002; 106: 1453-9.
- [109] Kolovou G, Anagnostopoulou K, Mikhailidis DP, Cokkinos DV. Apolipoprotein E knockout models. Curr Pharm Des 2008; 14: 338-51.
- [110] Ohsawa I, Nishimaki K, Yamagata K, Ishikawa M, Ohta S. Consumption of hydrogen water prevents atherosclerosis in apolipoprotein E knockout mice. Biochem Biophys Res Commun 2008; 377: 1195-8.
- [111] Furukawa S, Fujita T, Shimabukuro M, et al. Increased oxidative stress in obesity and its impact on metabolic syndrome. J Clin Invest 2004; 114: 1752-61.
- [112] Kamimura N, Nishimaki K, Ohsawa I, Ohta S. Molecular Hydrogen Improves Obesity and Diabetes by Inducing Hepatic FGF21 and Stimulating Energy Metabolism in db/db Mice. Obesity (Silver Spring) 2011. in press.
- [113] Yao X, Panichpisal K, Kurtzman N, Nugent K. Cisplatin nephrotoxicity: a review. Am J Med Sci 2007; 334: 115-24.
- [114] Kitamura A, Kobayashi S, Matsushita T, Fujinawa H, Murase K. Experimental verification of protective effect of hydrogen-rich water against cisplatin-induced nephrotoxicity in rats using dynamic contrast-enhanced CT. Br. J. Radiol 2010; 83: 509-14.
- [115] Itoh T, Fujita Y, Ito M, et al. Molecular hydrogen suppresses FcepsilonRI-mediated signal transduction and prevents degranulation of mast cells. Biochem Biophys Res Commun 2009; 389: 651-6.
- [116] Cardinal JS, Zhan J, Wang Y, et al. Oral hydrogen water prevents chronic allograft nephropathy in rats. Kidney Int 2010; 77: 101-9.
- [117] Sato Y, Kajiyama S, Amano A, et al. Hydrogen-rich pure water prevents superoxide formation in brain slices of vitamin C-depleted SMP30/GNL knockout mice. Biochem Biophys Res Commun 2008; 375: 346-50.
- [118] Lin Y, Kashio A, Sakamoto T, Suzukawa K, Kakigi A, Yamasoba T. Hydrogen in drinking water attenuates noise-induced hearing loss in guinea pigs. Neurosci Lett 2011; 487: 12-6.
- [119] Gu Y, Huang CS, Inoue T, Yamashita T, Ishida T, Kang KM, Nakao A. Drinking Hydrogen Water Ameliorated Cognitive Impairment in Senescence-Accelerated Mice. Journal of Clinical Biochemistry and Nutrition 2010; 46: 269-76.
- [120] Qian LR, Cao F, Cui JG, et al. The Potential Cardioprotective Effects of Hydrogen in Irradiated Mice. J. Radiat. Res. (Tokyo) 2010; 51: 741-7.
- [121] Cai JM, Kang ZM, Liu K, et al. Neuroprotective effects of hydrogen saline in neonatal hypoxia-ischemia rat model. Brain Res 2009; 1256: 129-37.
- [122] Li J, Wang C, Zhang JH, Cai JM, Cao YP, Sun XJ. Hydrogen-rich saline improves memory function in a rat model of amyloid-betainduced Alzheimer's disease by reduction of oxidative stress. Brain Res 2010; 1328: 152-61.
- [123] Chen C, Chen Q, Mao Y, et al. Hydrogen-rich saline protects against spinal cord injury in rats. Neurochem Res 2010; 35: 1111-8.
- [124] Liu Q, Shen WF, Sun HY, et al. Hydrogen-rich saline protects against liver injury in rats with obstructive jaundice. Liver Int 2010; 30: 958-68.
- [125] Mao YF, Zheng XF, Cai JM, et al. Hydrogen-rich saline reduces lung injury induced by intestinal ischemia/reperfusion in rats. Biochem Biophys Res Commun 2009; 381: 602-5.
- [126] Qian L, Cao F, Cui J, et al. The potential cardioprotective effects of hydrogen in irradiated mice. J Radiat Res (Tokyo) 2010; 51: 741-7.

- [127] Shingu C, Koga H, Hagiwara S, Matsumoto S, Goto K, Yokoi I, Noguchi T. Hydrogen-rich saline solution attenuates renal ischemia-reperfusion injury. J Anesth 2010; 24: 569-74.
- [128] Sun Q, Cai J, Liu S, Liu Y, Xu W, Tao H, Sun X. Hydrogen-rich saline provides protection against hyperoxic lung injury. J Surg Res 2011; 165: e43-9.
- [129] Zheng J, Liu K, Kang Z, et al. Saturated hydrogen saline protects the lung against oxygen toxicity. Undersea Hyperb Med 2010; 37: 185-92.
- [130] Zheng X, Mao Y, Cai J, et al. Hydrogen-rich saline protects against intestinal ischemia/reperfusion injury in rats. Free Radic Res 2009; 43: 478-84
- [131] Oharazawa H, Igarashi T, Yokota T, et al. Protection of the retina by rapid diffusion of hydrogen: administration of hydrogen-loaded eye drops in retinal ischemia-reperfusion injury. Invest Ophthalmol Vis Sci 2010; 51: 487-92.
- [132] Kubota M, Shimmura S, Kubota S, et al. Hydrogen and N-acetyl-L-cysteine rescue oxidative stress-induced angiogenesis in a mouse corneal alkali-burn model. Invest Ophthalmol Vis Sci 2011; 52: 427-33.
- [133] Nakabayashi M. Review of the ischemia hypothesis for ocular hypertension other than congenital glaucoma and closed-angle glaucoma. Ophthalmologica 2004; 218: 344-9.
- [134] Thauer RK, Jungermann K, Decker K. Energy conservation in chemotrophic anaerobic bacteria. Bacteriol Rev 1977; 41: 100-80.
- [135] Levitt MD. Production and excretion of hydrogen gas in man. N. Engl. J. Med 1969; 281: 122-7.
- [136] Chiasson JL, Josse RG, Gomis R, Hanefeld M, Karasik A, Laakso M. Acarbose treatment and the risk of cardiovascular disease and hypertension in patients with impaired glucose tolerance: the STOP-NIDDM trial. JAMA 2003; 290: 486-94.
- [137] Hanefeld M, Cagatay M, Petrowitsch T, Neuser D, Petzinna D, Rupp M. Acarbose reduces the risk for myocardial infarction in type 2 diabetic patients: meta-analysis of seven long-term studies. Eur. Heart J 2004; 25: 10-6.
- [138] Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). UK Prospective Diabetes Study (UKPDS) Group. Lancet 1998; 352: 837-53.
- [139] Suzuki Y, Sano M, Hayashida K, Ohsawa I, Ohta S, Fukuda K. Are the effects of alpha-glucosidase inhibitors on cardiovascular events related to elevated levels of hydrogen gas in the gastrointestinal tract? FEBS Lett 2009; 583: 2157-9.

Accepted: June 20, 2011

Received: May 12, 2011

- [140] Kajiya M, Sato K, Silva MJ, et al. Hydrogen from intestinal bacteria is protective for Concanavalin A-induced hepatitis. Biochem. Biophys. Res. Commun 2009; 386: 316-21.
- [141] Kajiya M, Silva MJ, Sato K, Ouhara K, Kawai T. Hydrogen mediates suppression of colon inflammation induced by dextran sodium sulfate. Biochem Biophys Res Commun 2009; 386: 11-5.
- [142] Shimouchi A, Nose K, Takaoka M, Hayashi H, Kondo T. Effect of dietary turmeric on breath hydrogen. Dig Dis Sci 2009; 54: 1725-9.
- [143] Chen X, Zuo Q, Hai Y, Sun XJ. Lactulose: an indirect antioxidant ameliorating inflammatory bowel disease by increasing hydrogen production. Med Hypotheses 2011; 76: 325-7.
- [144] Kajiyama S, Hasegawa G, Asano M, et al. Supplementation of hydrogen-rich water improves lipid and glucose metabolism in patients with type 2 diabetes or impaired glucose tolerance. Nutr Res 2008; 28: 137-43.
- [145] Nakao A, Toyoda Y, Sharma P, Evans M, Guthrie N. Effectiveness of hydrogen rich water on antioxidant status of subjects with potential metabolic syndrome-an open label pilot study. J Clin Biochem Nutr 2010; 46: 140-9.
- [146] Nakayama M, Nakano H, Hamada H, Itami N, Nakazawa R, Ito S. A novel bioactive haemodialysis system using dissolved dihydrogen (H2) produced by water electrolysis: a clinical trial. Nephrol. Dial. Transplant 2010; 25: 3026-33.
- [147] Xie K, Yu Y, Pei Y, Hou L, Chen S, Xiong L, Wang G. Protective effects of hydrogen gas on murine polymicrobial sepsis via reducing oxidative stress and HMGB1 release. Shock 2010; 34: 90-7.
- [148] Chen H, Sun YP, Li Y, et al. Hydrogen-rich saline ameliorates the severity of l-arginine-induced acute pancreatitis in rats. Biochem Biophys Res Commun 2010; 393: 308-13.
- [149] Sun Q, Kang Z, Cai J, et al. Hydrogen-rich saline protects myocardium against ischemia/reperfusion injury in rats. Exp Biol Med (Maywood) 2009; 234: 1212-9.
- [150] Jazwa A, Cuadrado A. Targeting heme oxygenase-1 for neuroprotection and neuroinflammation in neurodegenerative diseases. Curr Drug Targets 2010; 11: 1517-31.
- [151] Park DJ, Agarwal A, George JF. Heme oxygenase-1 expression in murine dendritic cell subpopulations: effect on CD8+ dendritic cell differentiation in vivo. Am J Pathol 2010; 176: 2831-9.
- [152] Ohta S, Nakao A, Ohno K. The 2011 Medical Molecular Hydrogen Symposium: An Inaugural Symposium of the Journal Medical Gas Research. Medical Gas Research 2011; 1: 10.